

Adaptive divergence and genetic vulnerability of relict species under climate change: a case study of *Pterocarya macroptera*

Tian-Rui Wang¹, Hong-Hu Meng^{2,✉}, Nian Wang^{3,✉}, Si-Si Zheng¹, Yun Jiang¹, Duo-Qing Lin¹,
Yi-Gang Song^{1,4,*} and Gregor Kozłowski^{1,4,5}

¹Eastern China Conservation Centre for Wild Endangered Plant Resources, Shanghai Chenshan Botanical Garden, Shanghai, 201602, China, ²Plant Phylogenetics and Conservation Group, Center for Integrative Conservation, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences, Kunming, 650223, China, ³State Forestry and Grassland Administration Key Laboratory of Silviculture in Downstream Areas of the Yellow River, College of Forestry, Shandong Agricultural University, Tai'an, 271018, China, ⁴Department of Biology and Botanic Garden, University of Fribourg, Fribourg, CH-1700, Switzerland and ⁵Natural History Museum Fribourg, Fribourg, CH-1700, Switzerland

*For correspondence. E-mail cherish-faith@163.com

Received: 16 May 2023 Returned for revision: 15 June 2023 Editorial decision: 23 June 2023 Accepted: 4 July 2023

• **Background and Aims** Understanding adaptive genetic variation and whether it can keep pace with predicted future climate change is critical in assessing the genetic vulnerability of species and developing conservation management strategies. The lack of information on adaptive genetic variation in relict species carrying abundant genetic resources hinders the assessment of genetic vulnerability. Using a landscape genomics approach, this study aimed to determine how adaptive genetic variation shapes population divergence and to predict the adaptive potential of *Pterocarya macroptera* (a vulnerable relict species in China) under future climate scenarios.

• **Methods** We applied restriction site-associated DNA sequencing (RAD-seq) to obtain 8244 single-nucleotide polymorphisms (SNPs) from 160 individuals across 28 populations. We examined the pattern of genetic diversity and divergence, and then identified outliers by genetic differentiation (F_{ST}) and genotype–environment association (GEA) methods. We further dissected the effect of geographical/environmental gradients on genetic variation. Finally, we predicted genetic vulnerability and adaptive risk under future climate scenarios.

• **Key Results** We identified three genetic lineages within *P. macroptera*: the Qinling-Daba-Tianmu Mountains (QDT), Western Sichuan (WS) and Northwest Yunnan (NWY) lineages, which showed significant signals of isolation by distance (IBD) and isolation by environment (IBE). IBD and IBE explained 3.7–5.7 and 8.6–12.8 % of the genetic structure, respectively. The identified GEA SNP-related genes were involved in chemical defence and gene regulation and may exhibit higher genetic variation to adapt to the environment. Gradient forest analysis revealed that the genetic variation was mainly shaped by temperature-related variables, indicating its adaptation to local thermal environments. A limited adaptive potential was suggested by the high levels of genetic vulnerability in marginal populations.

• **Conclusions** Environmental gradient mainly shaped the population differentiation of *P. macroptera*. Marginal populations may be at high risk of extinction, and thus proactive management measures, such as assisted gene flow, are required to ensure the survival of these populations.

Key words: Climate change, genetic offset, genotype–environment association, RAD-seq, relict species.

INTRODUCTION

Climate change potentially alters habitat suitability at the regional scale and results in local extinctions (Parmesan, 2006; Wiens, 2016), and has been considered to be a major driver of species range shifts and biodiversity loss (Scheffers *et al.*, 2016; Waldvogel *et al.*, 2020; Meng *et al.*, 2021). Species may survive during climate change by tracking conditions to which they are currently adapted, exhibiting considerable phenotypic plasticity, and evolving adaptation strategies to new environments (Davis and Shaw, 2001; Parmesan, 2006; Capblancq *et al.*, 2020; Waldvogel *et al.*, 2020). For tree species with a long generation time, local adaptation is the most significant strategy to cope with climate change (Dauphin *et al.*, 2020; Gougherty *et al.*, 2021;

Meng *et al.*, 2022). Thus, understanding and quantifying the adaptive potential of tree species could not only reveal how they would survive in the context of climate change, but also benefit conservation and management strategies to cope with global biodiversity loss (Razgour *et al.*, 2019; Waldvogel *et al.*, 2020).

Advances in landscape genomics have enabled the elucidation of the molecular genetic basis of the local adaptation of tree species (Li *et al.*, 2017; Sork, 2018). Genotype–environment association (GEA) approaches can identify loci involved in local adaptation. The Mantel test and redundancy analysis (RDA) can detect the role of geographical or environmental variables in shaping genetic structure (Rellstab *et al.*, 2015). Genetic offset, which is measured by the locally adaptive allelic variation, can assess the amount of change in genomic composition required

for a population to track future environmental conditions (Ellis *et al.*, 2012; Capblancq *et al.*, 2020). Therefore, landscape genomics offers powerful tools to detect adaptive genetic variation and predict the genetic vulnerability of species to climate change (Wang *et al.*, 2021; Feng and Du, 2022).

Trees are the main components of forest ecosystems (Brodribb *et al.*, 2020; Fazan *et al.*, 2020). Tree species usually occupy heterogeneous environments, resulting in local adaptation (Savolainen, 2011; Sork *et al.*, 2013; Capblancq *et al.*, 2020). Assessing the impacts of environmental factors on local adaptation for tree species could help forecast the health of forest ecosystems (Sork *et al.*, 2013; Sork, 2018). Relict species carry abundant genetic information related to environmental changes and function as storehouses of biodiversity (Woolbright *et al.*, 2014). For instance, relict species distributed in the Sino-Japanese and Sino-Himalayan Floristic Regions of East Asia exhibit high genetic diversity and contain valuable genetic resources (Qiu *et al.*, 2011, 2017; Tang *et al.*, 2018). However, these species are vulnerable to future climate changes due to genetic drift and limited gene exchange (Yannic *et al.*, 2014; Bay *et al.*, 2018; Cao *et al.*, 2020). At present, conservation management of relict species is mainly conducted from the perspective of genetic diversity (Chen *et al.*, 2015; Wei *et al.*, 2016; Wu *et al.*, 2020; Xu *et al.*, 2021). However, assessment of the genetic vulnerability and adaptation of relict species to future climate changes is more efficient and critical for the conservation and reforestation of these valuable resources (Capblancq *et al.*, 2020; Gougherty *et al.*, 2021). The rapid development of landscape genomics during the last decade has provided unprecedented opportunity to explore the genetic vulnerability of relict species.

Pterocarya macroptera is a vulnerable Cenozoic relict tree species in China that grows at an altitude of between 1100 and 3500 m (Lu *et al.*, 1999; Song *et al.*, 2019, 2020). It is a member of the family Juglandaceae and includes three varieties: var. *macroptera*, var. *insignis*, and var. *delavayi* (Lu *et al.*, 1999). This species spans from 98°E to 120°E and exhibits considerable morphological variations (Song *et al.*, 2020). With such a wide geographical range, whether environmental or geographical gradients shape the population differentiation of *P. macroptera*, to what extent population differentiation is influenced by environment or geography, and how populations would respond to future climate conditions remain poorly understood.

In this study, we sequenced 160 individuals of *P. macroptera* from 28 populations covering its entire distribution. Based on single-nucleotide polymorphisms (SNPs) obtained from restriction site-associated DNA sequencing (RAD-seq) data, we formulated the following objectives: (1) to infer population genetic differentiation and genetic diversity; (2) to quantify the contributions of environmental and geographical variables in shaping the spatial distributions of genetic variation; and (3) to assess the vulnerable populations with a mismatch between genotype and environment.

MATERIALS AND METHODS

Field sampling, RAD-seq library preparation and sequencing

Samples of healthy and mature leaves were collected from 28 *P. macroptera* populations representing its entire geographical range. Samples were dried and kept in silica gel. Genomic

DNA was extracted from tissue using a Plant Genomic DNA Kit (Tiangen, Beijing, China). A total of 160 individuals were selected for sequencing, with four to eight individuals per population (Supplementary Data Table S1). Samples were individually barcoded, and RAD libraries were prepared according to Baird *et al.* (2008). Genomic DNA was digested with the restriction enzyme *Taq*I in a 30-μL reaction. Then adapter (containing individual unique 4- to 10-bp barcodes) ligation was performed on 200 ng DNA. Ligated DNA was pooled, purified, and PCR-amplified by an ABI GeneAmp 9700. DNA fragments with sizes between 300 and 500 bp were selected based on AMPure XP bead-based size selection steps. RAD libraries were sequenced on an Illumina HiSeq™ platform using 150-bp paired-end reads at Major Bio Pharm Technology, Shanghai, China.

Data processing

The quality of the RAD data was assessed using FastQC (Andrews, 2010). Adapter sequences and low-quality bases (QC < 20) from the tail of each read were removed using Trimmomatic v0.36 (Bolger *et al.*, 2014). Then, reads with length ~30 bp were discarded. The data quality was evaluated by calculating the GC content and the Q30 values. The cleaned paired reads were demultiplexed and trimmed to 120 bp in length using a next-generation sequencing-uniform script provided by Major Bio Pharm Technology. Variants were reference-aligned from the short reads using the STACKS v2.6.1 pipeline (Catchen *et al.*, 2013; Davey *et al.*, 2013). Firstly, the reference genome (*Pterocarya stenoptera*; Zhang *et al.*, 2022) was indexed using BWA v0.7.17, and the paired-end reads of each sample were mapped to the reference genome based on the BWA-MEM algorithm (Li and Durbin, 2009). Next, the aligned reads were sorted by SAMTOOLS v1.15.1 (Li *et al.*, 2009), and the gstacks module in STACKS was used to create loci using a sliding window algorithm with default parameters. Finally, using the populations module in STACKS, the SNPs were obtained and filtered according to the following criteria: (1) markers with an observed heterozygosity > 0.70 among individuals were removed to reduce the potential occurrence of paralogues; (2) minor allele frequency (MAF) < 0.05; (3) only the first SNP locus of each read was retained by parameter `write_single_snp` to reduce physical linkage; and (4) a minimum of 70 % of individuals within a population were required to process a locus.

VCFtools v0.1.13 was used to discard loci with missing data present in at least 20 % of individuals and to keep only biallelic SNPs (Danecek *et al.*, 2011). Variant sites described in the VCF format were functionally annotated based on the *P. stenoptera* gene models using SnpEff v4.3t (Cingolani *et al.*, 2012). For SNPs, we performed an online BLAST search via the National Center for Biotechnology Information (NCBI) website based on the protein sequences of genes annotated by SnpEff. Based on the highest percentage identity of BLAST alignment, combined with the results of each gene identified by the UniProt database (<https://www.uniprot.org/>), we determined the possible molecular function of each gene and the biological process involved.

Population structure and genetic diversity

We investigated the maximum likelihood ancestor of each individual for all SNPs (obtained above) using ADMIXTURE v1.30 (Alexander *et al.*, 2009; Alexander and Lange, 2011).

For ADMIXTURE, the range of the co-ancestry clusters (K) was set to vary between 1 and 10. The optimal number of clusters was determined based on the lowest cross-validation error rate. For each K value, we performed a 10-fold cross-validation. The genetic differentiation among populations was calculated based on all SNPs using principal component analysis (PCA) in the R package *adeigenet* 2.1-5 (Jombart *et al.*, 2010; R Core Team, 2019). Population differentiation (F_{ST}) among lineages was calculated based on different SNP datasets using VCFtools v0.1.13 (Danecek *et al.*, 2011). We calculated the number of private alleles (N_p), percentage of polymorphic loci (PL), mean observed heterozygosity (H_o) and mean expected heterozygosity (H_e) using PLINK v1.9 with default settings based on all SNPs and all outlier SNPs (Purcell *et al.*, 2007). To minimize differences resulting from sample size variations, we also calculated unbiased allele richness for all SNPs and all outlier SNPs using the R package *hierfstat* 0.5-11 (Goudet, 2005; Halbert and Derr, 2008). The unbiased estimation of nucleotide diversity (π) was conducted using *pixy* v1.2.7 for all loci (including non-polymorphic loci) and all outlier loci following a window length of 10 kb (Korunes and Samuk, 2021). For each summary statistic, Tukey HSD tests were used to evaluate significant differences.

Environmental variables and their responses to genetic variation

Nineteen bioclimatic variables for the current period (1970–2000) at 30 arcsec resolution were downloaded from the WorldClim v.2.1 database based on the geographical coordinates of the sampled populations (Fick and Hijmans, 2017). Elevation was also included as it may be important to alpine species. We further downloaded predicted environmental variables in the defined period (2081–2100) using low- and high-emission scenarios of the shared socioeconomic pathway (SSP126 and SSP585; Meinshausen *et al.*, 2020). Gradient forest (GF) analysis was performed to identify 20 environmental variables that best explained the distribution of genetic variation using the R package *gradientForest* 0.1-17 based on all SNPs (Ellis *et al.*, 2012). Gradient forest models apply a non-parametric machine-learning regression algorithm tree to explore non-linear associations of spatial, environmental and allelic variables (Fitzpatrick and Keller, 2015). Two thousand regression trees were used for each SNP to fit the GF model, while keeping all parameters as default values. After evaluating the ranked accuracy importance among 20 environmental variables, the absolute value of Pearson correlation coefficients among these environmental variables was evaluated using the R package *usdm* 1.1-18 (Naimi *et al.*, 2014). Environmental variables across the range of *P. macroptera* with correlation coefficients $|r| < 0.7$ and the top three variables identified by the GF analysis were retained. Finally, eight variables (BIO02, mean diurnal range; BIO03, isothermality; BIO04, temperature seasonality; BIO07, temperature annual range; BIO08, mean temperature of wettest quarter; BIO13, precipitation of wettest month; BIO15, precipitation seasonality; and elevation) were used for subsequent analysis (Supplementary Data Tables S2 and S3). The eight environmental variables were used to build the final GF model, which was used to predict the genetic vulnerability of each population across the range of *P. macroptera*

(see section Prediction of genetic vulnerability). Given that the correlation between the top three variables identified by GF and the other variables may affect the final prediction of GF, we also used bioclimatic variables with correlation coefficients $|r| < 0.7$ (i.e. BIO02, BIO07, BIO08, BIO13 and BIO15) to build the final GF model.

Identification of selection signatures

The F_{ST} -based methods identified outlier SNPs as those with levels of differentiation above those of neutral SNPs among populations (Beaumont and Nichols, 1996). The GEA-based methods identified outlier SNPs as those exhibiting allele frequency changes that were strongly related to environmental variables (De Mita *et al.*, 2013). We focused on the F_{ST} -based (Whitlock and Lotterhos, 2015; Luu *et al.*, 2017) and GEA-based approaches (Günther and Coop, 2013; Caye *et al.*, 2019) to detect outlier SNPs that were subjected to selection.

The R packages PCADAPT 4.3.3 and OutFLANK 0.2 were employed as F_{ST} -based methods to identify F_{ST} outlier SNPs. These two methods were shown to have the lowest false discovery rate (FDR) (Luu *et al.*, 2017). PCADAPT identifies outlier SNPs with respect to population structure through PCA (Privé *et al.*, 2020). Three principal components ($K = 3$) captured most background genetic variation based on the results of ADMIXTURE and PCA (see section Genomic divergence and genetic diversity). The SNPs that deviated significantly from the neutral background structure along the principal components (Bonferroni correction with adjusted P -values < 0.05) were identified as putatively genetic differentiation loci. OutFLANK employs an improved likelihood approach to estimate the null distribution of population differentiation for neutral loci (Whitlock and Lotterhos, 2015). This program does not take demographic history into account and is less prone to false positives (Whitlock and Lotterhos, 2015). We used a left and right trim value of 0.05 for the null distribution of F_{ST} with default settings. SNPs with a minimum heterozygosity for loci > 0.1 and the desired FDR threshold < 0.05 were identified as putatively genetic differentiation loci.

Bayesian mixed modelling and mixed linear modelling were implemented in BAYENV (Coop *et al.*, 2010; Günther and Coop, 2013) combined with latent factor mixed modelling (LFMM; Caye *et al.*, 2019) to investigate the genomic basis of adaptation across contemporary climate gradients. A null hypothesis empirical model was established using putatively non- F_{ST} genetic loci (removed loci identified by PCADAPT and OutFLANK) in BAYENV. An allele frequency versus environment variable covariance matrix among populations was calculated with 10^6 iterations in five independent runs. Based on the prior null hypothesis distribution model, we calculated the posterior distribution P -values of the correlation between allele frequency and environmental variables. The Bayes factor (BF) was generated by calculating the correlation between the allele frequency and environmental variable covariance matrices after 10^5 runs. The BF was calculated five times, and the average BF was treated as the final matrix of the BF. SNPs with BF values > 10 and among the top 5 % with absolute value of Spearman rank correlation coefficients (ρ) were considered as significantly environment-associated loci. To reduce the

FDR, the associations between genotypes and environmental variables were determined using the LFMM method, which considers the population structure. Genetic data were converted into LFMM format using the R package LEA 2.0.0 (Frichot and François, 2015). Based on the clustering analysis results, $K = 3$ was selected as the optimal number of latent factors for sparse non-negative matrix factorization (SNMF). Missing SNP data were imputed based on the inferred genetic structure using the built-in SNMF function in the LEA package. Then, we ran ten independent operations to simulate the correlation between allele frequency and environmental variables for a burn-in period of 5,000 steps followed by 10,000 iterations. We used the `lfmm.pvalues` function in the LEA package to adjust the P -values. SNPs with adjusted P -values < 0.001 strongly support associations between allele frequencies and environmental variables. To verify the robust detection of outlier loci, we further corrected the P -values at an FDR of 0.05 using the R package `fdrtool` 1.2.17 (Strimmer, 2008).

Using the results of these four approaches, we divided our dataset into four categories for the subsequent analyses: (1) all SNPs; (2) putative F_{ST} SNPs detected jointly by PCADAPT and OutFLANK; (3) putative GEA SNPs detected jointly by BAYENV and LFMM; and (4) putative outlier SNPs identified by the F_{ST} and GEA methods. Finally, we used a Venn diagram to evaluate the consistency of outlier SNPs identified across the F_{ST} and GEA methods, respectively.

Inferring isolation by distance and isolation by environment

Isolation by distance (IBD) and isolation by environment (IBE) were inferred using the R package `vegan` 2.5.6 to investigate the role of geographical and environmental variables in shaping spatial genetic differentiation (Forester et al., 2018; Oksanen et al., 2019). The geographical distance (pairwise Euclidean difference, representing IBD) between sites was calculated using the R package `geosphere` 1.5.14 (Hijmans et al., 2021). The eight environmental variables were first scaled and centralized to account for differences in magnitude, and then were used to calculate the environmental distance (Bray–Curtis distance, representing IBE) using the R package `ecodist` 2.0.9 (Goslee and Urban, 2007). The genetic distance (pairwise $F_{ST}/(1 - F_{ST})$) was calculated based on all SNPs using the R package `hierfstat` 0.5.11 (Goudet, 2005). Simple Mantel tests (Mantel, 1967; Diniz-Filho et al., 2013) were used to test whether genetic distance was significantly correlated with geographical or environmental distance. Partial Mantel tests were used to investigate the independent effects of geographical/environmental distance on genetic distance. We further investigated the correlation between environmental and geographical distance. The level of significance of the Mantel tests was assessed with 999 permutations.

Redundancy analysis

We used RDA to assess the relative contribution of environmental and geographical distances to population genetic differentiation using the R package `vegan` 2.5-6 (Forester et al., 2018; Oksanen et al., 2019). Allele frequency matrices for each population, as well as two independent matrices of

environmental and geographical variables, were used for RDA tests. Considering that the RDA test does not allow for the presence of missing allele frequency data, we used the R package LEA 2.0.0 (Frichot and François, 2015) to impute missing allele frequencies for each population. Geographical variables were defined by geographical distances using principal coordinates of neighbourhood matrices (PCNMs) in the R package `vegan` (Oksanen et al., 2019). The first half of the PCNM variables (PCNM1, PCNM2, PCNM3 and PCNM4) with significant positive eigenvalues were retained, as has been suggested by Fitzpatrick and Keller (2015). Following the recommendation of Borcard et al. (2011), we used all SNPs to perform forward selection for both geographical and environmental variables with an α value of 0.05 to avoid overfitting. This resulted in the retention of three PCNMs (PCNM1, PCNM2 and PCNM3) and seven environmental variables (BIO02, BIO03, BIO04, BIO07, BIO13, BIO15 and elevation) for the subsequent analyses. Full and partial RDA model tests for different SNP sets (all and F_{ST} , GEA and all outlier SNPs) were performed to distinguish the independent effects of environment and geography by reciprocally constraining geographical and environmental variables. Significance was assessed using the randomization procedure implemented in the function `ANOVA.cca` with 999 randomizations.

Prediction of genetic vulnerability

To predict genetic vulnerability under future climate conditions, we performed GF (Ellis et al., 2012; Fitzpatrick and Keller, 2015) and risk of non-adaptedness analysis (RONA; Rellstab et al., 2016; Pina-Martins et al., 2018) using all SNPs and GEA SNPs. Gradient forest analysis was used to identify the spatial regions in which genotype–environment relationships are most likely to be disrupted by climate change. We evaluated the mismatch between current and predicted genomic compositions under future environmental projections during 2081–2100 under low- and high-emission scenarios (SSP126 and SSP585). Eight environmental variables were included in the GF model to predict the genomic composition of each grid point across the range of *P. macroptera*. The GF model was tested using 2000 regression trees per SNP. The Euclidian distance between current and future genetic compositions was calculated; this represents the scale of genetic change needed to match environmental change (i.e. genetic offset), with higher values indicating greater vulnerability of the population (Fitzpatrick and Keller, 2015). To illustrate the regions predicted to experience greater impacts under future environments with a lack of adaptive evolution or migration (Fitzpatrick and Keller, 2015), the genetic offset was visualized as landscape maps for all SNPs and GEA SNPs.

We used RONA to quantify the theoretical average change in allele frequency under predicted future climate scenarios and then predicted the adaptive potential of species under these scenarios. First, the allele frequency of each individual was obtained using the R package LEA 2.0.0 (Frichot and François, 2015). A regression model was constructed based on the allele frequencies and eight environmental variables. The theoretically expected allele frequencies during 2081–2100 under the SSP126 and SSP585 emission scenarios were then

predicted based on the regression model. The average difference between the current and predicted allele frequencies was the RONA value, which represented the adaptive potential of the population under future climate conditions. A higher RONA value indicates a lower potential (high genetic vulnerability) of the population to adapt to future climate conditions. We calculated the weighted mean R^2 of the regression model for each population, as recommended by Pina-Martins *et al.* (2018). Finally, we identified the top three environmental variables that were most tightly associated with all SNP and GEA SNP sets.

RESULTS

Genomic data

A total of 1 125 776 716 clean paired-end reads were obtained from 160 individuals of *P. macroptera*. An average of 7 036 104 reads was retained per individual. The average mapping rate was ~77 % (range 60–94 %; Supplementary Data Tables S1 and S4). We retained 8244 high-quality SNPs after stringent quality control (Supplementary Data Table S5). The annotation of the 8244 SNPs is provided in Supplementary Data Table S6. Of the

8244 SNPs, 1779 (21.6 %) resided in coding regions. The remaining SNPs (6465 SNPs, 78.4 %) resided in upstream gene variants (1812 SNPs), downstream gene variants (1257 SNPs), intron variants (859 SNPs) and intergenic regions (2537 SNPs; Supplementary Data Table S6).

Genomic divergence and genetic diversity

Based on the 8244 SNPs, ADMIXTURE identified $K = 3$ as the most likely number of evolutionary clusters among the 28 populations (Fig. 1A, B; Supplementary Data Fig. S1). We detected three distinct clusters: the Qinling-Daba-Tianmu Mountain (QDT), Western Sichuan (WS) and Northwest Yunnan (NWY) lineages. PCA yielded a similar grouping, with the first two PCs accounting for 20.8 % of the total genetic variation using 8244 SNPs (16.2 and 4.6 % for PC1 and PC2 respectively; Fig. 1C). The genetic differentiation based on the different SNP datasets consistently supported the highest genetic differentiation between the QDT and NWY lineages, followed by the NWY and WS lineages, while the QDT and WS lineages had the lowest genetic differentiation (Table 1). In addition, four SNP datasets revealed different levels of genetic differentiation. Among them, the differentiation level based on F_{ST}

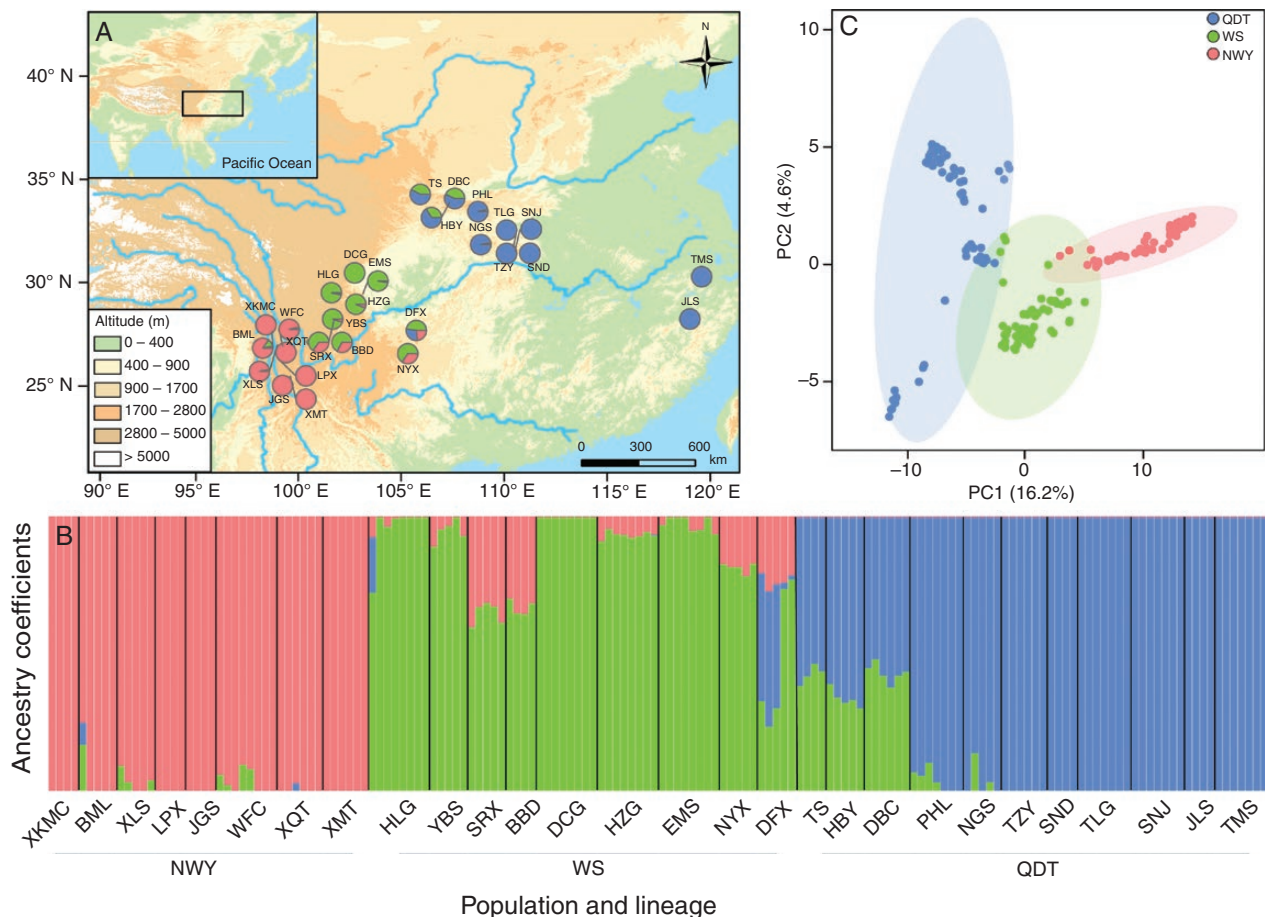


FIG. 1. Geographical distribution and population genomic structure in *P. macroptera*. (A) Mapping of the Bayesian genetic clusters inferred by ADMIXTURE. Colours represent different ancestry groups. (B) Ancestry assignment for 160 individuals in 28 populations of *P. macroptera* at $K = 3$. Each bar represents an individual, with different colours reflecting varying ancestry. (C) PCA with different colours reflecting different groups.

TABLE 1. Genetic differentiation (F_{ST}) on all 8244 SNPs and all outlier SNPs (in parentheses, below the diagonal) and on F_{ST} SNPs (PCADAPT and OutFLANK, 932 SNPs) and GEA SNPs (BAYENV and LFMM, 957 SNPs, in parentheses, above the diagonal) between groups of *P. macroptera*.

	NWY lineage	WS lineage	QDT lineage
NWY lineage	—	0.478 (0.382)	0.656 (0.609)
WS lineage	0.155 (0.383)	—	0.323 (0.288)
QDT lineage	0.281 (0.575)	0.084 (0.249)	—

SNPs was 0.32–0.66, which was higher than that based on GEA SNPs (0.29–0.61) and all outlier SNPs (0.25–0.58). Based on all SNPs, the genetic differentiation among different lineages was the lowest (0.09–0.28). For all SNPs, the genetic diversity indicated no significant difference among the three lineages ($P < 0.05$; Table 2; Supplementary Data Tables S7 and S8). For all outlier SNPs, the WS lineage showed more polymorphic loci and higher allelic richness (AR) than the NWY and QDT lineages. Specifically, the HLG population in the WS lineage had the greatest proportion of polymorphic loci (53.49 %) and the DFX population had the highest allelic richness (1.39) among the 28 populations (Supplementary Data Table S7).

Putative outlier SNP detection

A total of 932 outlier SNPs were detected using the F_{ST} -based methods, while 537 and 697 outlier SNPs were identified by PCADAPT and OutFLANK, respectively (Table 3; Supplementary Data Table S9; Supplementary Data Fig. S2). The number of top 20 % outlier SNPs identified by PCADAPT was 117, and that by OutFLANK was 139, with 17 outlier SNPs overlapping between the two methods (Supplementary Data Fig. S3; Supplementary Data Table S10). Among these 17 overlapping outlier SNPs, ten were located in ten genes and the remaining seven SNPs were located in intergenic regions. Of the ten genes, the PST000227 gene was not identified by the NCBI BLAST analysis and the PST000586 gene had no known function. Three genes (PST026568, PST019941 and PST000475) were involved in embryo development ending in seed dormancy, the triterpenoid biosynthetic process and the reproductive development process (Supplementary Data Table S10).

Based on the GEA approach, a total of 957 outlier SNPs putatively associated with environmental variables were detected. Among them, 431 and 796 SNPs were detected by BAYENV and LFMM ($FDR < 0.05$), respectively (Table 3; Supplementary Data Table S11; Supplementary Data Figs S4 and S5). We found 356, 306, 290, 288 and 283 SNPs associated with the environmental variables BIO13, BIO07, BIO04, BIO03 and BIO15, respectively. BAYENV and LFMM detected 49 and 47 SNPs simultaneously associated with the top four environmental variables (BIO04, BIO03, BIO07, and BIO15; see next section) (Supplementary Data Fig. S6A, B). Eight SNPs were detected by both BAYENV and LFMM (Supplementary Data Fig. S6C). Among these eight SNPs, one was located in an intergenic region, and the remaining seven SNPs were

TABLE 2. Genetic diversity for *P. macroptera* under all SNPs and all outlier SNPs.

Lineage	N	N_p	PL	H_o	H_e	π	Allelic richness
All SNPs							
NWY	5.25 ^a	0.00 ^a	33.54 ^a	0.11 ^a	0.10 ^a	0.12 ^a	1.27 ^a
WS	6.22 ^a	0.33 ^a	45.16 ^a	0.12 ^a	0.11 ^a	0.13 ^a	1.31 ^a
QDT	5.63 ^a	0.27 ^a	37.08 ^a	0.11 ^a	0.10 ^a	0.11 ^a	1.27 ^a
All outlier SNPs							
NWY	5.25 ^a	0.00 ^a	30.98 ^b	0.09 ^a	0.10 ^{ab}	0.12 ^{ab}	1.26 ^b
WS	6.22 ^a	0.11 ^a	43.64 ^a	0.12 ^a	0.13 ^a	0.15 ^a	1.33 ^a
QDT	5.63 ^a	0.09 ^a	30.00 ^b	0.09 ^a	0.09 ^b	0.10 ^b	1.24 ^b

N , number of individuals; N_p , number of private alleles; PL, percentage of polymorphic loci; H_o , mean observed heterozygosity; H_e , mean expected heterozygosity; π , mean nucleotide diversity.

Indices with different superscript letters represent significant differences ($P < 0.05$, Tukey–HSD).

located in seven genes, respectively. Of the seven genes, the PST000586 and PST036282 genes had no known function. The PST020689 and PST029083 genes were involved in the regulation of gene expression by stress factors and photosynthesis, respectively. In addition, the PST029941 gene was involved in the glycolipid and sulfolipid biosynthetic process, and the PST018122 gene was responsible for the diterpenoid, sesquiterpene and terpenoid biosynthetic process (Supplementary Data Table S10). A total of 16 genes were detected by the F_{ST} and GEA approaches, among which the PST035198 gene, involved in the regulation of gene expression, was detected by both methods.

Environmental and spatial associations with genetic variation

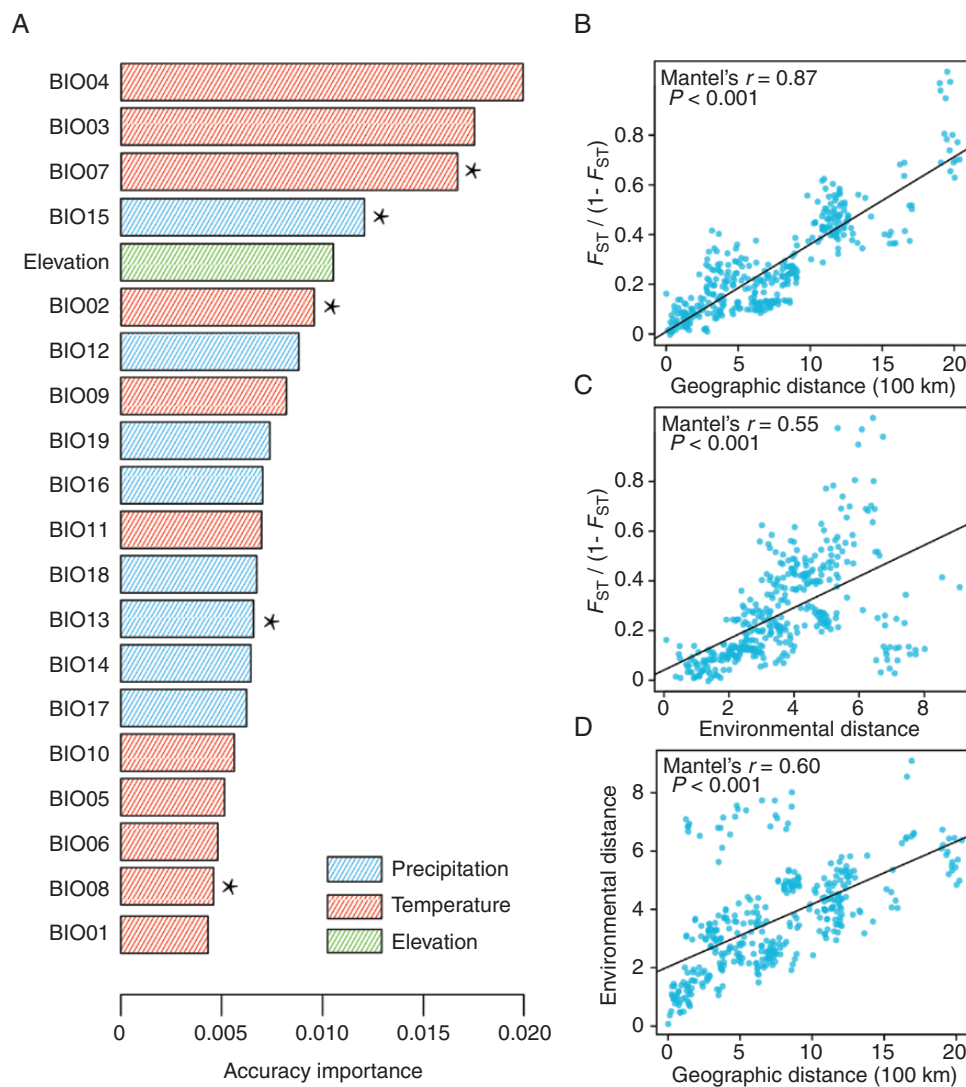
The GF analysis revealed significant differences in genetic composition along the geographical range of *P. macroptera* (Fig. 2A; Supplementary Data Table S12). Temperature seasonality (BIO04) was identified as the most important predictor among the environmental variables considered, followed by BIO03, BIO07, BIO15 and elevation. In addition, allelic composition changed sharply along the temperature-related top three environmental variables, BIO04, BIO03, and BIO07 (Supplementary Data Fig. S7).

Using all SNPs, the pairwise genetic distance between locations [$F_{ST}/(1 - F_{ST})$] was highly correlated with geographical distance, suggesting a strong signal of IBD (Fig. 2B). Genetic distance was also significantly associated with environmental distance based on eight environmental variables (Fig. 2C). In a partial Mantel test, only geographical distance was significant (Supplementary Data Table S13), suggesting that this is the strongest force structuring genome-wide variation. In addition, the autocorrelation between environmental and geographical distances was also strong (Fig. 2D).

Isolation by distance explained 3.7–5.7 % of the variation after controlling for environment, while IBE explained 8.6–12.8 % of the variation after controlling for geography (Table 4; Supplementary Data Table S14). Based on the four SNP

TABLE 3. Number of candidate SNP loci under putative selection identified by PCADAPT, OutFLANK, BAYENV and LFMM.

Method		SNPs	BIO02	BIO03	BIO04	BIO07	BIO08	BIO13	BIO15	Elevation
F_{ST} -based	PCADAPT	537								
	OutFLANK	697								
	F_{ST} total	932								
GEA-based	BAYENV	431	80	207	166	183	8	28	124	20
	LFMM	796	102	167	186	201	132	353	222	149
	GEA Total	957	162	288	290	306	133	356	283	157
	All outlier SNPs ^a	1380								

^aSNPs identified by F_{ST} - and GEA-based methods.FIG. 2. (A) Ranked importance of environmental variables based on GF modelling. The asterisks represent the uncorrelated environment variables (Pearson's $|r| < 0.7$). (B–D) Mantel tests of genetic distance [$F_{ST} / (1 - F_{ST})$] against (B) geographical distance and (C) environmental distance, and (D) correlation between geographical distance and environmental distance.

datasets, the contribution of environment to genetic variation was higher than that of geography. Thus, we further identified the explanatory environmental variables for the genetic variation

in different lineages of *P. macroptera* using different SNP sets. Considering the similar results of RDA and partial RDA, we reported RDA results with a higher proportion of explained

TABLE 4. Summary of genetic differentiation associated with environmental, geographical and their combined effects based on redundancy analysis in *P. macroptera*.

	All SNPs 8244	F_{ST} SNPs 932	GEA SNPs 957	All outlier SNPs 1380
Combined fractions				
F~geog.	0.200***	0.481***	0.448***	0.415***
F~env.	0.249***	0.545***	0.533***	0.484***
Individual fractions				
F~geog. env.	0.037***	0.057***	0.042***	0.050***
F~env. geog.	0.086***	0.121***	0.128***	0.119***
Total explained	0.286***	0.601***	0.576***	0.534***
Total confounded	0.163	0.423	0.406	0.365
Total unexplained	0.714	0.399	0.424	0.466
Total	1.000	1.000	1.000	1.000

F, dependent matrix of minor allele frequencies; RDA tests are of the form: F ~ independent matrices | covariate matrices. env., seven retained environmental variables; geog., three retained principal coordinates of neighbourhood matrices variables. Total explained, total adjusted R^2 of individual fractions; Total confounded, total of individual fractions confounded between various combinations of environment and geography.

*** $P \leq 0.001$.

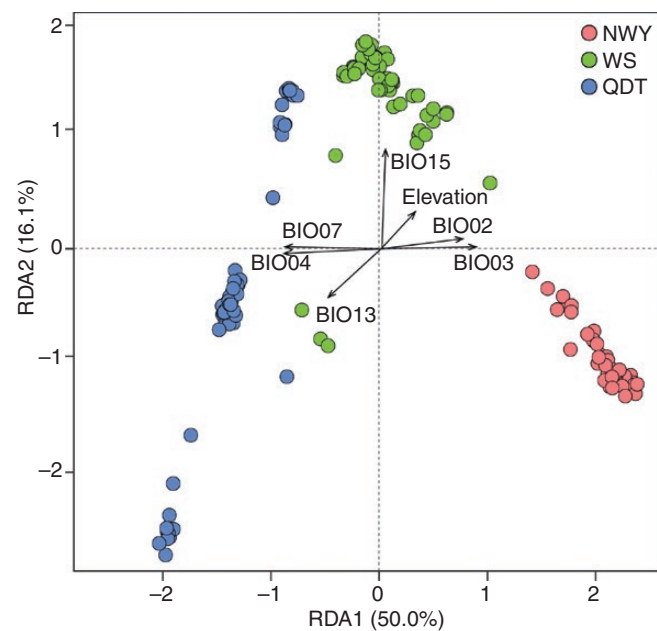


FIG. 3. Partial RDA partitioning sources of genetic differentiation among populations in *P. macroptera* into environment by condition geography using all outlier SNPs. The plot shows the first and second RDA axes with individuals as coloured circles and environmental variables as black vectors.

genetic variation. Based on the four SNP datasets, the contribution of environmental variables to genetic variation in the three lineages was generally consistent (Fig. 3; Supplementary Data Fig. S8). Precipitation-related variables (BIO15, BIO13) and elevation contributed most to the genetic variation in the WS lineage. Temperature-related variables (BIO04, BIO07) explained most of the genetic variation in the QDT lineage. Mean diurnal range (BIO02) and isothermality (BIO03) contributed most to the genetic variation in the NWY lineage. A total of

28.6–60.1 % of the variation was explained by two components (environment and geography) of the different SNP sets (‘Total explained’ in Table 4), and a large proportion of total genetic variation was explained by their combined effect (‘Total confounded’ in Table 4). This combined effect was most pronounced in the F_{ST} SNPs.

Landscape of future predictions

Gradient forest prediction based on five environmental variables was similar to those based on the eight environment variables (Supplementary Data Fig. S9). Given the similarity of the results, we reported predictions based on the eight environment variables. The ranges and degree of genetic mismatch increased under SSP585 compared with those under SSP126 based on all SNPs and GEA SNPs. Comparing the two scenarios, we calculated the proportion of the distribution range having a genetic mismatch > 50 % of the maximum detected value (0.22 in this study; Fig. 4). There was no distribution space exceeding the threshold of vulnerability based on all SNPs. Based on GEA SNPs, 3.4 and 6.9 % of the distribution space was recognized as above the threshold of vulnerability under SSP126 and SSP585, respectively. The eastern and western peripheral populations of *P. macroptera* were most vulnerable under both climate scenarios. This indicates that these populations may be confronted with climate-induced selective pressure in the future. The northern populations of the QDT lineage and the eastern populations of the NWY lineage had low genetic vulnerability (Fig. 4).

The RONA suggested that GEA SNPs had a higher genetic vulnerability than all SNPs (Fig. 5; Supplementary Data Table S15). Most populations under the high-emission scenario had a higher genetic vulnerability than those under the low-emission scenario. Precipitation of wettest month (BIO13), temperature seasonality (BIO04) and precipitation seasonality (BIO15) played a primary role in vulnerability. The southernmost and

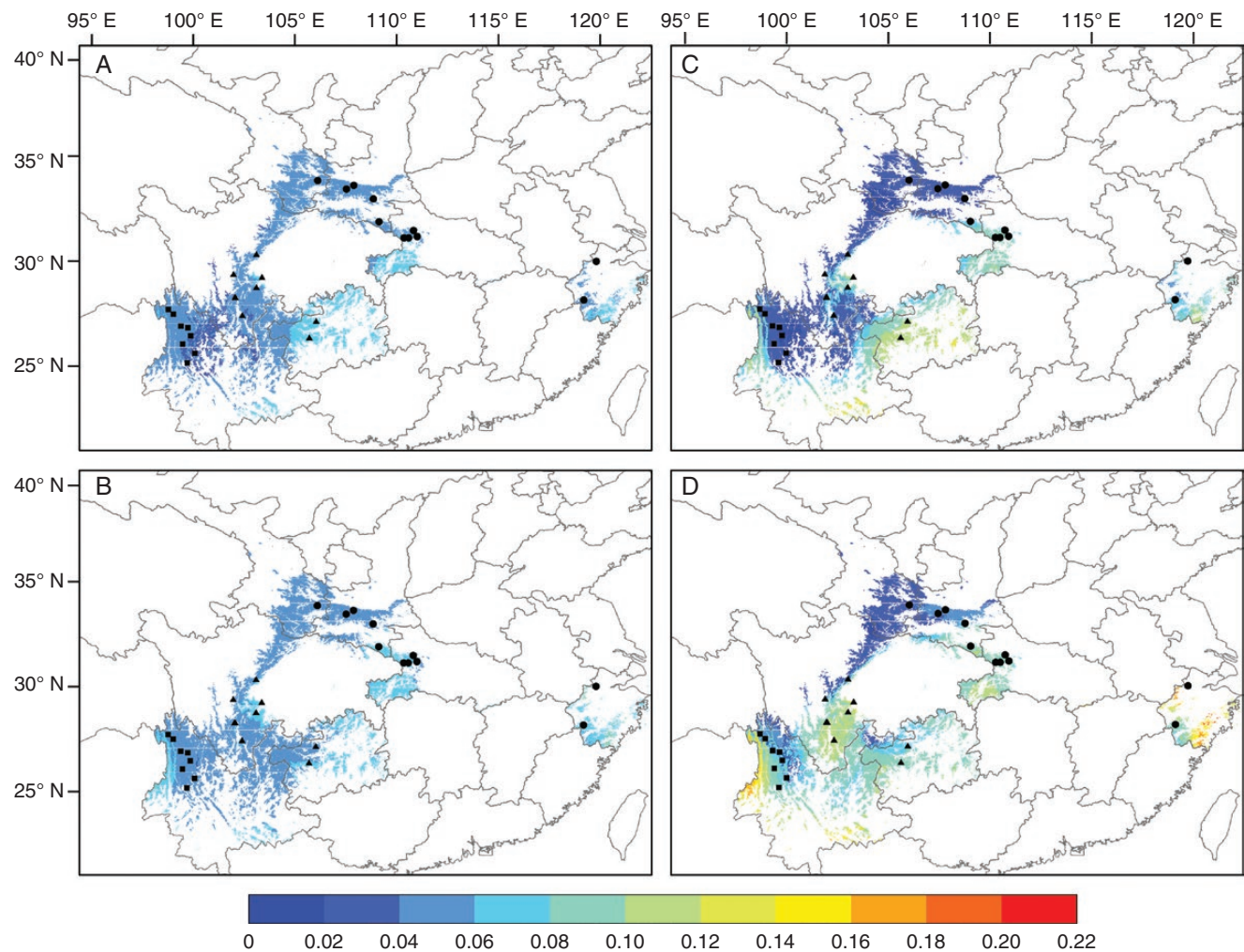


FIG. 4. Prediction of genetic offset to future climate change based on eight environment variables for (A, B) all SNPs and (C, D) GEA SNPs. (A) and (C) reflect scenario SSP126 2081–2100; (B) and (D) reflect scenario SSP585 2081–2100. Colour scale at bottom represents genomic offset. Black dots represent sampling sites on the geographical map, with squares, triangles and circles representing the Northwest Yunnan (NWY), Western Sichuan (WS) and Qinling-Daba-Tianmu Mountain (QDT) lineage, respectively.

easternmost marginal populations of *P. macroptera*, such as the populations JGS, DFX, NYX, TMS and JLS, had a lower adaptation potential for BIO13. Populations isolated at the southern and eastern edge of their natural distribution area may face higher vulnerability under predicted future climate scenarios.

DISCUSSION

Genomic divergence and diversity

We found three genetic clusters within *P. macroptera* with some admixed individuals between them. Two genetic clusters (NWY and WS) were located in the Sino-Himalayan Forest subkingdom and one (QDT) in the Sino-Japanese Forest subkingdom. This pattern of population structure is consistent with previous studies in East Asia (Wu and Wu, 1996; Qiu *et al.*, 2011, 2017; Chen *et al.*, 2015; Ma *et al.*, 2015; Cao *et al.*, 2016, 2020; Wei *et al.*, 2016). The two genetic clusters within the Sino-Himalayan Forest subkingdom seem to mirror the

similar phylogeographic break in many other species (Meng *et al.*, 2015; Yang *et al.*, 2017; Luo *et al.*, 2018; Wang *et al.*, 2019; Nocchi *et al.*, 2023). The level of genetic differentiation among *P. macroptera* populations is higher than that in other closely related wingnut taxa (*P. stenoptera*: 0.067) (Li *et al.*, 2018). The high genetic differentiation between the three lineages within *P. macroptera* may be related to the geographical isolation and divergent selection. Some SNPs located in different lineages may be subject to divergent selection in the homogenous gene pool. The WS lineage, mainly located in the west of the Sichuan Basin, has a higher level of genetic diversity, possibly due to the presence of an ancient refugium, which provided stable and suitable conditions for maintenance of genetic diversity (López-Pujol *et al.*, 2011; Tang *et al.*, 2018). Previous reports on plants (Li *et al.*, 2023), birds (Wu *et al.*, 2017), amphibians (Qiao *et al.*, 2018) and insects (Tang *et al.*, 2022) have shown that mountains around the Sichuan Basin harboured suitable microenvironments for species, especially for relict species. Another possible explanation for the high genetic diversity of the WS lineage could be the mixed populations in this lineage

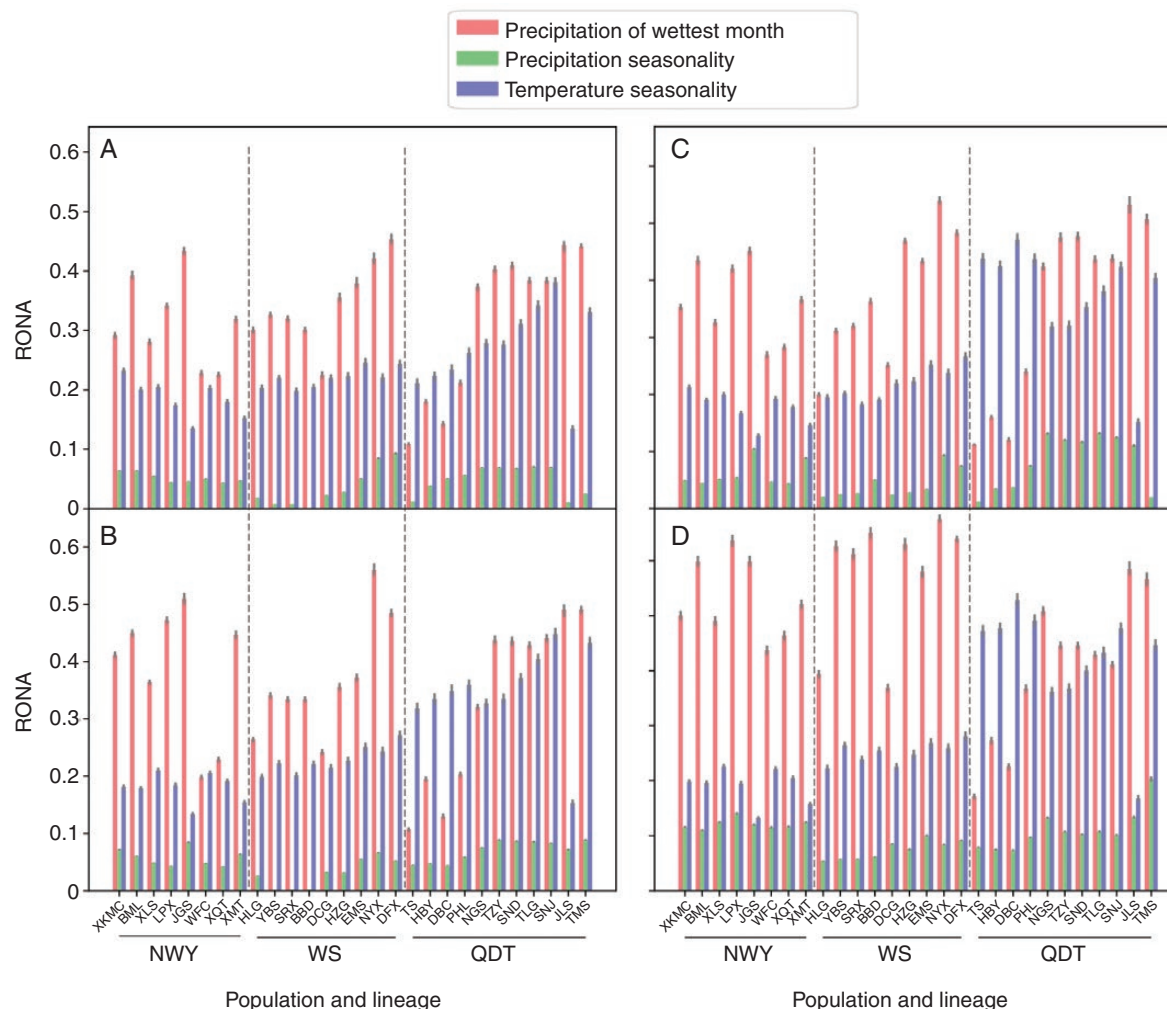


FIG. 5. RONA plot for the three most associated environmental variables for (A, B) all SNPs and (C, D) GEA SNPs. (A) and (C) reflect scenario SSP126 2081–2100; (B) and (D) reflect scenario SSP585 2081–2100. Bars represent the weighted means (by R^2 value) and lines at the tops of bars represent the standard error.

(i.e. SRX, BBD, BYX, DFX) receiving genetic introgression from the NWY and QDT lineages and thus providing higher genetic diversity for the WS lineage. The low genetic diversity in the QDT and NWY lineages requires further work to investigate the evolutionary dynamics of extinction, colonization and effective population size in these two lineages.

Landscape of current adaptive genetic variation

Trees often occupy highly heterogeneous environments and ecologically adapt to the local environment (Capblancq *et al.*, 2020). Hence, the signature of environmental isolation of *P. macroptera* likely reflects local environmental adaptation (Nachman and Payseur, 2012; Wang and Bradburd, 2014). Temperature regimes and precipitation patterns are critical factors that define species distribution and plant growth (Root *et al.*, 2003; Wahid *et al.*, 2007). Redundancy analysis showed that temperature-related factors were important in accounting for the adaptive variation of *P. macroptera*. This is similar to other findings that temperature also impacts the adaptive genetic variation of *P. stenoptera* (Li *et al.*, 2018, 2022), a species closely related to *P. macroptera*. We are not sure if

the same subsets of loci are involved in the local adaptation of both *P. macroptera* and *P. stenoptera*. Further research is needed to determine this. The top three environmental variables explaining genetic variation were temperature-related, especially for the NWY and QDT lineages. *Pterocarya* is a typical riparian relict tree genus (Song *et al.*, 2020, 2021). Compared with other *Pterocarya* species, *P. macroptera* occupies the highest elevations, with the largest change in elevational range, along mountain streams and water-rich slopes (Kozłowski *et al.*, 2018; Song *et al.*, 2019). This is likely why the other two important factors that drove genetic variation in *P. macroptera* were precipitation seasonality and elevation, which were also the top two factors for the WS lineage. The uplift of the Qinghai–Tibet Plateau created elevation gradients in East Asia, and intensification of the East Asian monsoon system shaped temperature regimes and precipitation patterns (Qiu *et al.*, 2011, 2017), which are the most suggestive factors responsible for genetic variation and the major phylogeographic breaks among the three lineages.

The GEA-based methods identified seven SNPs located in seven genes that were associated with environmental variables (BIO04, BIO03, BIO07, and BIO15). The identified

GEA genes involved in chemical defence and gene regulation may exhibit high genetic variation to adapt to the environment (temperature and precipitation). The PST018122 and PST019941 genes (detected by the F_{ST} method) were both involved in the triterpenoid biosynthetic process. As significant chemical defence compounds for the growth and development of plants and for coping with a stressed environment, terpenoids directly act as antimicrobials or signals for resisting herbivores and other natural enemies (Dudareva *et al.*, 2004; Paschold *et al.*, 2006). Terpenoids dominate in the leaves of *Pterocarya* species (Xiao *et al.*, 2002; Liu *et al.*, 2004; Zhang *et al.*, 2006; Ebrahimzadeh *et al.*, 2009; Yin *et al.*, 2019), suggesting that terpene biosynthesis-related genes may play an important role in protecting the leaves of *P. macroptera*. The PST020689 gene was involved in the regulation of gene expression by stress factors. Regulation of biological processes is responsible for stress resistance under biotic and abiotic stresses (Licausi *et al.*, 2013). Genes related to stress resistance may be of significance to *P. macroptera* under stressed conditions. The regulatory expression of these identified genes may be the genomic imprint for the local adaptation of *P. macroptera* to its habitats.

Genetic vulnerability under future climate conditions

Understanding the genetic basis of adaptation and determining the adaptive ability of species to future climate conditions are crucial in the context of climate change (Fitzpatrick and Keller, 2015; Dauphin *et al.*, 2020). Previous studies reported relevant strategies to gain insight into the potential risk of species persistence under climate change (Bay *et al.*, 2018; Du *et al.*, 2020; Zhao *et al.*, 2020; Vranken *et al.*, 2021; Sang *et al.*, 2022). We found that marginal populations of *P. macroptera* had a high level of genetic vulnerability, suggesting that these populations are potentially at higher risk of *in situ* extinction under future climate changes. These marginal populations may be less resilient to future climates because genotypes were not sufficiently correlated with predicted climate variables. This result reinforces our understanding that ecologically marginal populations may be separated not only by distance from the core of the species' distribution but also experience different biotic and abiotic environments (Munwes *et al.*, 2010; Santini *et al.*, 2019). Although the static elevation variable used for the prediction could have led to an underestimation of genetic vulnerability, riparian forest niches limited the movement of populations to higher elevations. In addition, the strong geographical isolation limited the ability of populations to track spatial changes through migration.

The genetic vulnerability of *P. macroptera* predicted in this study may be related to the species-specific tolerance to environmental variables and the complex topography of mountains. Overall, we expect that long-term sustained climate change will result in marginal populations with high genetic vulnerability (Dauphin *et al.*, 2020). These highly vulnerable populations need to adapt quickly to climate change. Otherwise they may be at risk of extinction (Franks and Hoffmann, 2012; Capblancq *et al.*, 2020). Future work needs to integrate adaptive genetic variation with biogeographic

models to accurately identify species vulnerability under future climate conditions.

Implications for conservation management

There is a growing interest in evolutionarily informed management strategies that rely on the spatial distribution of genetic diversity and genetic vulnerability (Aitken and Whitlock, 2013; Lefèvre *et al.*, 2014; Gougherty *et al.*, 2021). Hence, a comprehensive understanding of the spatial genetic diversity helps to develop conservation strategies (Petit *et al.*, 1998). Populations with high genetic diversity may have a greater potential in adaption to climate change and may harbour valuable breeding materials (Frankel *et al.*, 1995). Therefore, priority should be given to conservation of such populations, such as populations of the WS lineage of *P. macroptera*.

Gradient forest analysis and RONA have been widely used in assessing genetic vulnerability. Our results suggested that marginal populations of *P. macroptera* have higher genetic vulnerability. Assisted gene flow from populations with genotypes preadapted to future climate may help those marginal populations mitigate future climate change (Kremer *et al.*, 2012; Aitken and Bemmels, 2016). Thus, mixing seeds from multiple sources may be an appropriate strategy for increasing diversity and buffering climate change for marginal populations of *P. macroptera* (Aitken and Bemmels, 2016; Martins *et al.*, 2018). Meanwhile, donor populations should be carefully selected, so that transplanted individuals are genetically compatible with the new environment predicted in future reconstructive management (Fredriksen *et al.*, 2020).

SUPPLEMENTARY DATA

Supplementary data are available online at [https://academic.oup.com/aob](https://academic.oup.com/aob/advance-article/doi/10.1093/aob/mcab083/7220463) and consist of the following.

Figure S1: ADMIXTURE bar plots of the proportion of genetic membership for each ancestry. Figure S2: F_{ST} outlier SNPs in *P. macroptera* identified by PCADAPT and OutFLANK. Figure S3: unique and shared outlier SNPs for the top 20 % F_{ST} SNPs identified by PCADAPT and OutFLANK. Figure S4: Manhattan plot of SNPs called by BAYENV in *P. macroptera* with eight environment variables. Figure S5: Manhattan plot of SNPs called by LFMM in *P. macroptera* with eight environmental variables. Figure S6: unique and shared outlier SNPs associated with the top four environmental variables. Figure S7: cumulative importance of genetic variation along environmental gradients in *P. macroptera*. Figure S8: partial redundancy analysis by condition geography using all SNPs, F_{ST} SNPs identified in PCADAPT and OutFLANK, and GEA SNPs identified in BAYENV and LFMM. Figure S9: prediction of genetic offset to future climate change based on five environment variables for all SNPs and GEA SNPs. Table S1: summary of statistical information on sequencing quality for 28 populations of *P. macroptera*. Table S2: details of population locations, sample size and current environmental variables for 28 populations of *P. macroptera*. Table S3: predicted environmental variables for the years 2081–2100 under two shared socio-economic pathways for *P. macroptera*. Table S4: genetic information statistics on mapping rate and missing rate of 160 individuals in 28

populations. Table S5: statistical information for 8244 SNPs. Table S6: functional annotation of 8244 SNPs based SnpEff software. Table S7: genetic diversity for 28 populations of *P. macroptera*. Table S8: unbiased estimation of nucleotide diversity for NWY, WS and QDT lineages of *P. macroptera*. Table S9: outlier SNPs detected by F_{ST} -based methods. Table S10: functional descriptions of genes associated with SNPs identified by F_{ST} - and GEA-based methods. Table S11: outlier SNPs detected by GEA-based methods. Table S12: accuracy importance of each environmental variable identified by GF modelling for *P. macroptera*. Table S13: partial Mantel test in *P. macroptera* conditioned with environmental and geographical distance. Table S14: partitioning of the variance and accumulated constrained eigenvalues associated with environment based on partial redundancy analysis for all SNPs, F_{ST} SNPs, GEA SNPs, and all outlier SNPs. Table S15: summary of risk of non-adaptedness calculated for SSP126 and SSP585 in *P. macroptera* populations based on future climate predictions for 2081–2100.

FUNDING

This work was supported by the Special Fund for Scientific Research of Shanghai Landscaping & City Appearance Administrative Bureau (G212406, G202401), Foundation Franklinia, the Yunnan Revitalization Talent Support Program in Yunnan Province (H.H.M.) and the CAS Light of West China Program (H.H.M.).

ACKNOWLEDGEMENTS

We want to thank Mr Rui-Bin Liu for helping with material collection. We would like to thank Editage (www.editage.cn) for English language editing. Y.G.S. and G.K. designed the study; S.S.Z., Y.G.S., Y.J. and D.Q.L. performed the material collections and experiments; T.R.W. performed the simulations and the genomic analysis; T.R.W., H.H.M., N.W., Y.G.S., and G.K. wrote the manuscript; all authors read and approved the manuscript. The data underlying this article are available at Figshare: <https://doi.org/10.6084/m9.figshare.23742066>.

LITERATURE CITED

- Aitken SN, Bemmels JB. 2016. Time to get moving: assisted gene flow of forest trees. *Evolutionary Applications* 9: 271–290. doi:10.1111/eva.12293.
- Aitken SN, Whitlock MC. 2013. Assisted gene flow to facilitate local adaptation to climate change. *Annual Review of Ecology, Evolution, and Systematics* 44: 367–388. doi:10.1146/annurev-ecolsys-110512-135747.
- Alexander DH, Lange K. 2011. Enhancements to the ADMIXTURE algorithm for individual ancestry estimation. *BMC Bioinformatics* 12: 246.
- Alexander DH, Novembre J, Lange K. 2009. Fast model-based estimation of ancestry in unrelated individuals. *Genome Research* 19: 1655–1664. doi:10.1101/gr.094052.109.
- Andrews S. 2010. *Fast QC: A quality control tool for high throughput sequence data*. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/> (3 January 2023, date last accessed).
- Baird NA, Etter PD, Atwood TS, et al. 2008. Rapid SNP discovery and genetic mapping using sequenced RAD markers. *PLoS One* 3: e3376. doi:10.1371/journal.pone.0003376.
- Bay RA, Harrigan RJ, Underwood VL, Gibbs HL, Smith TB, Ruegg K. 2018. Genomic signals of selection predict climate-driven population declines in a migratory bird. *Science* 359: 83–86. doi:10.1126/science.aan4380.
- Beaumont MA, Nichols RA. 1996. Evaluating loci for use in the genetic analysis of population structure. *Proceedings of the Royal Society of London. Series B: Biological Sciences* 263: 1619–1626.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30: 2114–2120. doi:10.1093/bioinformatics/btu170.
- Borcard D, Gillet F, Legendre P. 2011. *Numerical ecology with R*. New York: Springer.
- Brodrick TJ, Powers J, Cochard H, Choat B. 2020. Hanging by a thread? Forests and drought. *Science* 368: 261–266. doi:10.1126/science.aat7631.
- Cao YN, Comes HP, Sakaguchi S, Chen LY, Qiu YX. 2016. Evolution of East Asia's Arcto-Tertiary relict *Euptelea* (Eupteleaceae) shaped by late Neogene vicariance and Quaternary climate change. *BMC Evolutionary Biology* 16: 1–17.
- Cao YN, Zhu SS, Chen J, et al. 2020. Genomic insights into historical population dynamics, local adaptation, and climate change vulnerability of the East Asian Tertiary relict *Euptelea* (Eupteleaceae). *Evolutionary Applications* 13: 2038–2055. doi:10.1111/eva.12960.
- Capblancq T, Fitzpatrick MC, Bay RA, Exposito-Alonso M, Keller SR. 2020. Genomic prediction of (mal)adaptation across current and future climatic landscapes. *Annual Review of Ecology, Evolution, and Systematics* 51: 245–269. doi:10.1146/annurev-ecolsys-020720-042553.
- Catchen J, Hohenlohe PA, Bassham S, Amores A, Cresko WA. 2013. Stacks: an analysis tool set for population genomics. *Molecular Ecology* 22: 3124–3140. doi:10.1111/mec.12354.
- Caye K, Jumentier B, Lepeule J, François O. 2019. LFMM 2: fast and accurate inference of gene-environment associations in genome-wide studies. *Molecular Biology and Evolution* 36: 852–860.
- Cingolani P, Platts A, Wang LL, et al. 2012. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff. *Fly* 6: 80–92. doi:10.4161/fly.19695.
- Chen JM, Zhao SY, Liao YY, Gichira AW, Gituru RW, Wang QF. 2015. Chloroplast DNA phylogeographic analysis reveals significant spatial genetic structure of the relictual tree *Davidia involucrata* (Davidiaceae). *Conservation Genetics* 16: 583–593.
- Coop G, Witonsky DB, Rienzo AD, Pritchard JK. 2010. Using environmental correlations to identify loci underlying local adaptation. *Genetics* 185: 1411–1423.
- Danecek P, Auton A, Abecasis G, et al.; 1000 Genomes Project Analysis Group. 2011. The variant call format and VCFtools. *Bioinformatics* 27: 2156–2158. doi:10.1093/bioinformatics/btr330.
- Dauphin B, Rellstab C, Schmid MZS, et al. 2020. Genomic vulnerability to rapid climate warming in a tree species with a long generation time. *Global Change Biology* 27: 1181–1195. doi:10.1111/gcb.15469.
- Davey JW, Cezard T, Fuentes-Utrilla P, Eland C, Gharbi K, Blaxter ML. 2013. Special features of RAD sequencing data: implications for genotyping. *Molecular Ecology* 22: 3151–3164. doi:10.1111/mec.12084.
- Davis MB, Shaw RG. 2001. Range shifts and adaptive responses to Quaternary climate change. *Science* 292: 673–679. doi:10.1126/science.292.5517.673.
- Diniz-Filho JAF, Soares TN, Lima JS, et al. 2013. Mantel test in population genetics. *Genetics and Molecular Biology* 36: 475–485. doi:10.1590/S1415-47572013000400002.
- Dudareva N, Pichersky E, Gershenzon J. 2004. Biochemistry of plant volatiles. *Plant Physiology* 135: 1893–1902. doi:10.1104/pp.104.049981.
- Du FK, Wang TR, Wang YY, Ueno S, de Lafontaine G. 2020. Contrasted patterns of local adaptation to climate change across the range of an evergreen oak, *Quercus aquifolioides*. *Evolutionary Applications* 13: 2377–2391. doi:10.1111/eva.13030.
- Ebrahimzadeh MA, Nabavi SF, Nabavi SM. 2009. Essential oil composition and antioxidant activity of *Pterocarya fraxinifolia*. *Pakistan Journal of Biological Sciences* 12: 957–963.
- Ellis N, Smith SJ, Pitcher, CR. 2012. Gradient forests: calculating importance gradients on physical predictors. *Ecology* 93: 156–168.
- Fazan L, Song YG, Kozłowski G. 2020. The woody planet: from past triumph to manmade decline. *Plants* 9: 1593. doi:10.3390/plants9111593.
- Feng L, Du FK. 2022. Landscape genomics in tree conservation under a changing environment. *Frontiers in Plant Science* 13: 822217.
- Fick SE, Hijmans RJ. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* 37: 4302–4315. doi:10.1002/joc.5086.
- Fitzpatrick MC, Keller SR. 2015. Ecological genomics meets community-level modelling of biodiversity: mapping the genomic landscape of

- current and future environmental adaptation. *Ecology Letters* **18**: 1–16. doi:10.1111/ele.12376.
- Forester BR, Lasky JR, Wagner HH, Urban DL. 2018. Comparing methods for detecting multilocus adaptation with multivariate genotype–environment associations. *Molecular Ecology* **27**: 2215–2233. doi:10.1111/mec.14584.
- Frankel OH, Brown AHD, Burdon J. 1995. The genetic diversity of wild plants. In: *The conservation of plant biodiversity*. ?Impact-insert-start?>Cambridge: Cambridge University Press, 10–38.
- Franks SJ, Hoffmann AA. 2012. Genetics of climate change adaptation. *Annual Review of Genetics* **46**: 185–208. doi:10.1146/annurev-genet-110711-155511.
- Fredriksen S, Filbee-Dexter K, Norderhaug KM, et al. 2020. Green gravel: a novel restoration tool to combat kelp forest decline. *Scientific Reports* **10**: 3983. doi:10.1038/s41598-020-60553-x.
- Frichot E, François O. 2015. LEA: an R package for landscape and ecological association studies. *Methods in Ecology and Evolution* **6**: 925–929. doi:10.1111/2041-210X.12382.
- Goslee SC, Urban DL. 2007. The ecodist package for dissimilarity-based analysis of ecological data. *Journal of Statistical Software* **22**: 1–19.
- Goudet J. 2005. HIERFSTAT, a package for R to compute and test hierarchical *F*-statistics. *Molecular Ecology Notes* **5**: 184–186. doi:10.1111/j.1471-8286.2004.00828.x.
- Gougherty AV, Keller SR, Fitzpatrick MC. 2021. Maladaptation, migration and extirpation fuel climate change risk in a forest tree species. *Nature Climate Change* **11**: 166–171. doi:10.1038/s41558-020-00968-6.
- Günther T, Coop G. 2013. Robust identification of local adaptation from allele frequencies. *Genetics* **195**: 205–220. doi:10.1534/genetics.113.152462.
- Halbert ND, Derr JN. 2008. Patterns of genetic variation in US federal bison herds. *Molecular Ecology* **17**: 4963–4977. doi:10.1111/j.1365-294X.2008.03973.x.
- Hijmans RJ, Williams E, Vennes C, Hijmans MR. 2021. Package ‘geosphere’. R package version 1.5-14. <https://cran.r-project.org/web/packages/geosphere/index.html> (1 April 2022, date last accessed).
- Jombart T, Devillard S, Balloux F. 2010. Discriminant analysis of principal components: a new method for the analysis of genetically structured populations. *BMC Genetics* **11**: 94. doi:10.1186/1471-2156-11-94.
- Korunes KL, Samuk K. 2021. pixy: unbiased estimation of nucleotide diversity and divergence in the presence of missing data. *Molecular Ecology Resources* **21**: 1359–1368. doi:10.1111/1755-0998.13326.
- Kozłowski G, Bétrisey S, Song YG. 2018. *Wingnuts (Pterocarya) and walnut family. Relict trees: linking the past, present and future*. Fribourg: Natural History Museum Fribourg.
- Kremer A, Ronce O, Robledo-Arnuncio JJ, et al. 2012. Long-distance gene flow and adaptation of forest trees to rapid climate change. *Ecology Letters* **15**: 378–392. doi:10.1111/j.1461-0248.2012.01746.x.
- Lefèvre F, Boivin T, Bontemps A, et al. 2014. Considering evolutionary processes in adaptive forestry. *Annals of Forest Science* **71**: 723–739.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* **25**: 1754–1760. doi:10.1093/bioinformatics/btp324.
- Li H, Handsaker B, Wysoker A, et al.; 1000 Genome Project Data Processing Subgroup. 2009. The sequence alignment/map format and SAMtools. *Bioinformatics* **25**: 2078–2079. doi:10.1093/bioinformatics/btp352.
- Li Y, Zhang XX, Mao RL, et al. 2017. Ten years of landscape genomics: challenges and opportunities. *Frontiers in Plant Science* **8**: 2136.
- Li JX, Zhu XH, Li Y, et al. 2018. Adaptive genetic differentiation in *Pterocarya stenoptera* (Juglandaceae) driven by multiple environmental variables were revealed by landscape genomics. *BMC Plant Biology* **18**: 306.
- Li LF, Cushman SA, He YX, et al. 2022. Landscape genomics reveals genetic evidence of local adaptation in a widespread tree, the Chinese wingnut (*Pterocarya stenoptera*). *Journal of Systematics and Evolution* **60**: 386–397.
- Li X, Ruhsam M, Wang Y, et al. 2023. Wind-dispersed seeds blur phylogeographic breaks: the complex evolutionary history of *Populus lasiocarpa* around the Sichuan Basin. *Plant Diversity* **45**: 156–168. doi:10.1016/j.pld.2022.10.003.
- Licausi F, Ohme-Takagi M, Perata P. 2013. APETALA 2/ethylene responsive factor (AP2/ERF) transcription factors: mediators of stress responses and developmental programs. *New Phytologist* **199**: 639–649.
- Liu HB, Cui CB, Cai B, et al. 2004. Isolation, identification and antitumor activity of triterpenes from *Pterocarya tonkinensis* (Franch.) Dode. *Chinese Journal of Medicinal Chemistry* **14**: 165–168.
- López-Pujol J, Zhang FM, Sun HQ, Ying TS, Ge S. 2011. Centres of plant endemism in China: places for survival or for speciation? *Journal of Biogeography* **38**: 1267–1280. doi:10.1111/j.1365-2699.2011.02504.x.
- Lu AM, Stone D E, Grauke LJ. 1999. Juglandaceae. In: Hong DY, Pan KY, Turland NJ, eds. *Flora of China*, Vol. 4. Beijing and Saint Louis: Science Press and Missouri Botanical Garden Press, 277–285.
- Luo D, Xu B, Rana SK, Li ZM, Sun H. 2018. Phylogeography of rare fern *Polystichum glaciale* endemic to the subnival zone of the Sino-Himalaya. *Plant Systematics and Evolution* **304**: 485–499. doi:10.1007/s00606-018-1495-2.
- Luu K, Bazin E, Blum MGB. 2017. pcadapt: an R package to perform genome scans for selection based on principal component analysis. *Molecular Ecology Resources* **17**: 67–77. doi:10.1111/1755-0998.12592.
- Ma Q, Du YJ, Chen N, Zhang LY, Li JH, Fu CX. 2015. Phylogeography of *Davidia involucreata* (Davidiaceae) inferred from cpDNA haplotypes and nSSR data. *Systematic Botany* **40**: 796–810.
- Mantel N. 1967. The detection of disease clustering and a generalized regression approach. *Cancer Research* **27**: 209–220.
- Martins K, Gugger PF, Llanderal-Mendoza J, et al. 2018. Landscape genomics provides evidence of climate-associated genetic variation in Mexican populations of *Quercus rugosa*. *Evolutionary Applications* **11**: 1842–1858. doi:10.1111/eva.12684.
- Meinshausen M, Nicholls ZR, Lewis J, et al. 2020. The shared socio-economic pathway (SSP) greenhouse gas concentrations and their extensions to 2500. *Geoscientific Model Development* **13**: 3571–3605.
- Meng LH, Chen G, Li ZH, Yang YP, Wang ZK, Wang LY. 2015. Refugial isolation and range expansions drive the genetic structure of *Oxyria sinensis* (Polygonaceae) in the Himalaya-Hengduan Mountains. *Scientific Reports* **5**: 10396.
- Meng HH, Gao XY, Song YG, Cao GL, Li J. 2021. Biodiversity arks in the Anthropocene. *Regional Sustainability* **2**: 109–115. doi:10.1016/j.regsus.2021.03.001.
- Meng HH, Zhang CY, Song YG, et al. 2022. Opening a door to the spatiotemporal history of plants from the tropical Indochina Peninsula to subtropical China. *Molecular Phylogenetics and Evolution* **171**: 107458. doi:10.1016/j.ympev.2022.107458.
- De Mita S, Thuillet AC, Gay L, et al. 2013. Detecting selection along environmental gradients: analysis of eight methods and their effectiveness for outbreeding and selfing populations. *Molecular Ecology* **22**: 1383–1399. doi:10.1111/mec.12182.
- Munwes I, Geffen E, Roll U, et al. 2010. The change in genetic diversity down the core-edge gradient in the eastern spadefoot toad (*Pelobates syriacus*). *Molecular Ecology* **19**: 2675–2689. doi:10.1111/j.1365-294X.2010.04712.x.
- Nachman MW, Payseur BA. 2012. Recombination rate variation and speciation: theoretical predictions and empirical results from rabbits and mice. *Philosophical Transactions of the Royal Society B: Biological Sciences* **367**: 409–421. doi:10.1098/rstb.2011.0249.
- Naimi B, Hamm NAS, Groen TA, Skidmore AK, Toxopeus AG. 2014. Where is positional uncertainty a problem for species distribution modelling? *Ecography* **37**: 191–203.
- Nocchi G, Wang J, Yang L, et al. 2023. Genomic signals of local adaptation and hybridization in Asian white birch. *Molecular Ecology* **32**: 595–612. doi:10.1111/mec.16788.
- Oksanen J, Blanchet FG, Kindt R, et al. 2019. *Vegan: Community Ecology Package*. R package version 2.5-6. <https://cran.r-project.org/web/packages/vegan/index.html> (11 October 2022, date last accessed).
- Parmesan C. 2006. Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology, Evolution, and Systematics* **37**: 637–669. doi:10.1146/annurev.ecolsys.37.091305.110100.
- Paschold A, Halitschke R, Baldwin IT. 2006. Using ‘mute’ plants to translate volatile signals. *Plant Journal* **45**: 275–291. doi:10.1111/j.1365-3113x.2005.02623.x.
- Petit RJ, Mousadik A, Pons O. 1998. Identifying populations for conservation on the basis of genetic markers. *Biological Conservation* **12**: 844–855.
- Pina-Martins F, Baptista J, Pappas JG, Paulo OS. 2018. New insights into adaptation and population structure of cork oak using genotyping by sequencing. *Global Change Biology* **25**: 337–350.
- Privé F, Luu K, Vilhjálmsson BJ, Blum MGB. 2020. Performing highly efficient genome scans for local adaptation with R package pcadapt version 4. *Molecular Biology and Evolution* **37**: 2153–2154. doi:10.1093/molbev/msaa053.

- Purcell S, Neale B, Todd-Brown K, et al. 2007. PLINK: A tool set for whole-genome association and population-based linkage analyses. *American Journal of Human Genetics* 81: 559–575. doi:10.1086/519795.
- Qiao L, Wen GN, Qi Y, et al. 2018. Evolutionary melting pots and reproductive isolation: a ring-shaped diversification of an odorous frog (*Odorrana margaritacea*) around the Sichuan Basin. *Molecular Ecology* 27: 4888–4900.
- Qiu YX, Fu CX, Comes HP. 2011. Plant molecular phylogeography in China and adjacent regions: tracing the genetic imprints of Quaternary climate and environmental change in the world's most diverse temperate flora. *Molecular Phylogenetics and Evolution* 59: 225–244. doi:10.1016/j.ympev.2011.01.012.
- Qiu YX, Lu QX, Zhang YH, Cao YN. 2017. Phylogeography of East Asia's Tertiary relict plants: current progress and future prospects. *Biodiversity Science* 25: 24–28. doi:10.17520/biods.2016292.
- R Core Team 2019. *R: a language and environment for statistical computing*. Vienna: R Foundation for Statistical Computing. <https://www.R-project.org/>.
- Razgour O, Forester B, Taggart JB, et al. 2019. Considering adaptive genetic variation in climate change vulnerability assessment reduces species range loss projections. *Proceedings of the National Academy of Sciences of the USA* 116: 10418–10423. doi:10.1073/pnas.1820663116.
- Reilstab C, Gugerli F, Eckert AJ, Hancock AM, Holderegger R. 2015. A practical guide to environmental association analysis in landscape genomics. *Molecular Ecology* 24: 4348–4370. doi:10.1111/mec.13322.
- Reilstab C, Zoller S, Walthert L, et al. 2016. Signatures of local adaptation in candidate genes of oaks (*Quercus* spp.) with respect to present and future climatic conditions. *Molecular Ecology* 25: 5907–5924. doi:10.1111/mec.13889.
- Root TL, Price JT, Hall KR, Schneider SH, Rosenzweig C, Pounds JA. 2003. Fingerprints of global warming on wild animals and plants. *Nature* 421: 57–60. doi:10.1038/nature01333.
- Sang YP, Long ZQ, Dan XM, et al. 2022. Genomic insights into local adaptation and future climate-induced vulnerability of a keystone forest tree species in East Asia. *Nature Communications* 13: 6541.
- Santini L, Pironon S, Maiorano L, Thuiller W. 2019. Addressing common pitfalls does not provide more support to geographical and ecological abundant-centre hypotheses. *Ecography* 42: 692–705.
- Savolainen O. 2011. The genomic basis of local climatic adaptation. *Science* 334: 49–50. doi:10.1126/science.1213788.
- Scheffers BR, De Meester L, Bridge TCL, et al. 2016. The broad footprint of climate change from genes to biomes to people. *Science* 354: aaf7671. doi:10.1126/science.aaf7671.
- Song YG, Bétrisey S, Kozłowski G. 2019. *Pterocarya macroptera*. The IUCN Red List of Threatened Species: e.T66816136A66816223.
- Song YG, Li Y, Meng HH, et al. 2020. Phylogeny, taxonomy, and biogeography of *Pterocarya* (Juglandaceae). *Plants* 9: 1524. doi:10.3390/plants9111524.
- Song YG, Walas L, Pietras M, et al. 2021. Past, present and future suitable areas for the relict tree *Pterocarya fraxinifolia* (Juglandaceae): integrating fossil records, niche modeling, and phylogeography for conservation. *European Journal of Forest Research* 140: 1323–1339. doi:10.1007/s10342-021-01397-6.
- Sork VL. 2018. Genomic studies of local adaptation in natural plant populations. *Journal of Heredity* 109: 3–15.
- Sork VL, Aitken SN, Dyer RJ, Eckert AJ, Legendre P, Neale DB. 2013. Putting the landscape into the genomics of trees: approaches for understanding local adaptation and population responses to changing climate. *Tree Genetics and Genomes* 9: 901–911. doi:10.1007/s11295-013-0596-x.
- Strimmer K. 2008. fdrtool: a versatile R package for estimating local and tail area-based false discovery rates. *Bioinformatics* 24: 1461–1462. doi:10.1093/bioinformatics/btn209.
- Tang CQ, Matsui T, Ohashi H, et al. 2018. Identifying long-term stable refugia for relict plant species in East Asia. *Nature Communications* 9: 4488.
- Tang XT, Lu MX, Du YZ. 2022. Molecular phylogeography and evolutionary history of the pink rice borer (Lepidoptera: Noctuidae): implications for refugia identification and pest management. *Systematic Entomology* 47: 371–383. doi:10.1111/syen.12535.
- Vranken S, Wernberg T, Scheben A, et al. 2021. Genotype-environment mismatch of kelp forests under climate change. *Molecular Ecology* 30: 3730–3746. doi:10.1111/mec.15993.
- Wahid A, Gelani S, Ashraf M, Foolad MR. 2007. Heat tolerance in plants: an overview. *Environmental and Experimental Botany* 61: 199–223. doi:10.1016/j.envexpbot.2007.05.011.
- Waldvogel AM, Feldmeyer B, Rolshausen G, et al. 2020. Evolutionary genomics can improve prediction of species' responses to climate change. *Evolution Letters* 4: 4–18. doi:10.1002/evl3.154.
- Wang IJ, Bradburd GS. 2014. Isolation by environment. *Molecular Ecology* 23: 5649–5662. doi:10.1111/mec.12938.
- Wang TR, Feng L, Du FK. 2021. New approaches for ecological adaptation study: from population genetics to landscape genomics. *Scientia Sinica Vitae* 51: 167–178.
- Wang ZW, Zhang TC, Luo D, Sun WG, Sun H. 2019. Phylogeography of *Excoecaria acerifolia* (Euphorbiaceae) suggests combined effects of historical drainage reorganization events and climatic changes on riparian plants in the Sino-Himalayan region. *Botanical Journal of the Linnean Society* 192: 350–368.
- Wei XZ, Sork VL, Meng HJ, Jiang MX. 2016. Genetic evidence for central-marginal hypothesis in a Cenozoic relict tree species across its distribution in China. *Journal of Biogeography* 43: 2173–2185. doi:10.1111/jbi.12788.
- Whitlock MC, Lotterhos KE. 2015. Reliable detection of loci responsible for local adaptation: inference of a null model through trimming the distribution of F_{ST} . *American Naturalist* 186: S24–S36. doi:10.1086/682949.
- Wiens JJ. 2016. Climate-related local extinctions are already widespread among plant and animal species. *PLoS Biology* 14: e2001104. doi:10.1371/journal.pbio.2001104.
- Woolbright SA, Whitham TG, Gehring CA, Allan GJ, Bailey JK. 2014. Climate relicts and their associated communities as natural ecology and evolution laboratories. *Trends in Ecology and Evolution* 29: 406–416. doi:10.1016/j.tree.2014.05.003.
- Wu ZY, Wu SG. 1996. A proposal for a new floristic kingdom (realm) – the E. Asiatic kingdom, its delimitation and characteristics. In: Zhang AL, Wu S, eds. *Proceedings of the First International Symposium on Floristic Characteristics and Diversity of East Asian Plants*. Beijing and Berlin: China Higher Education Press and Springer, 3–42.
- Wu YJ, DuBay SG, Colwell RK, Ran JH, Lei FM. 2017. Mobile hotspots and refugia of avian diversity in the mountains of south-west China under past and contemporary global climate change. *Journal of Biogeography* 44: 615–626.
- Wu XT, Ruhsam M, Wen YF, et al. 2020. The last primary forests of the Tertiary relict *Glyptostrobus pensilis* contain the highest genetic diversity. *Forestry* 93: 359–375.
- Xiao C, Gregg PC, Hu WL, Yang ZH, Zhang ZN. 2002. Attraction of the cotton bollworm, *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae), to volatiles from wilted leaves of a non-host plant, *Pterocarya stenoptera*. *Applied Entomology and Zoology* 37: 1–6. doi:10.1303/aez.2002.1.
- Xu WQ, Comes HP, Feng Y, Zhang YH, Qiu YX. 2021. A test of the centre-periphery hypothesis using population genetics in an East Asian Tertiary relict tree. *Journal of Biogeography* 48: 2853–2864. doi:10.1111/jbi.14244.
- Yang LQ, Hu HY, Xie C, et al. 2017. Molecular phylogeny, biogeography and ecological niche modelling of *Cardiocrinum* (Liliaceae): insights into the evolutionary history of endemic genera distributed across the Sino-Japanese floristic region. *Annals of Botany* 119: 59–72. doi:10.1093/aob/mcw210.
- Yannic G, Pellissier L, Ortego J, et al. 2014. Genetic diversity in caribou linked to past and future climate change. *Nature Climate Change* 4: 132–137.
- Yin CP, Sun FF, Rao Q, Zhang YL. 2019. Chemical compositions and antimicrobial activities of the essential oil from *Pterocarya stenoptera* C. DC. *Natural Product Research* 34: 2828–2831. doi:10.1080/14786419.2019.1587426.
- Zhang ZF, Wu WJ, Gao ZZ. 2006. Analysis of the volatile components of fresh, wilts, dry China wingnut (*Pterocarya stenoptera* C. DC.) by SPME with GC-MS. *Natural Product Research* 18: 778–783.
- Zhang WP, Cao L, Lin XR, et al. 2022. Dead-end hybridization in walnut trees revealed by large-scale genomic sequence data. *Molecular Biology and Evolution* 39: msab308. doi:10.1093/molbev/msab308.
- Zhao W, Sun YQ, Pan J, et al. 2020. Effects of landscapes and range expansion on population structure and local adaptation. *New Phytologist* 228: 330–343. doi:10.1111/nph.16619.